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The Tree Fern Highland Lace is a Cultivar of Sphaeropteris cooperi

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ABSTRACT.—The tree fern Highland Lace had an unusual introduction into cultivation almost thirty years ago in Eastern Australia and was initially identified as *Sphaeropteris tomentosissima* (Copel.) R.M.Tryon. Since then, it has been introduced to Europe and the US, and it remains a popular tree fern found in both public and private collections. We re-examined this fern, comparing it to a herbarium type specimen, and conclude that it is not *S. tomentosissima*, but is most likely a variant form of *Sphaeropteris cooperi* (F.V. Mueller) R.M.Tryon. Sequence analysis of chloroplast DNA [*rbcL*, *atpA* and *trnL* (UAA) intron] confirmed this species identification.

KEY WORDS.—Highland Lace, Sphaeropteris tomentosissima, Sphaeropteris cooperi, Sphaeropteris excelsa, tree ferns, chloroplast DNA sequence analysis, rbcL, atpA, trnL (UAA) intron

A distinctive tree fern with narrow pinnules and relatively small fronds appeared in Australian cultivation in the 1980s. It was a robust grower and its reduced pinnules imparted a lacy look to the leaves. Compared to most tree ferns, it was smaller, but it also seemed to bear more leaves in its crown. It originated as an unknown contaminant in a sporing pot at a wholesale nursery on the north coast of New South Wales, Australia. The late Rod Hill, an Australian tree fern enthusiast, made an attempt to identify the species and his closest match was Sphaeropteris tomentosissima (Copel.) R.M. Tryon, which grows in the highlands of west central New Guinea. In the 1980s this plant spread among tree fern collectors and commercial growers in Australia and by the 1990s it was being grown in Europe and the United States. It is called either Highland Lace, New Guinea Treefern or Sphaeropteris tomentosissima. Its lacier appearance compared to other cultivated tree ferns has led to its high popularity, and it was awarded a first place and a trophy at the Los Angeles International Fern Society's annual Exotic Plant Show in 1997 and 2003 (Lois and Kurt Rossten, Huntington Beach, California). Despite the enthusiasm for this new addition to the limited list of commercially available cultivated tree ferns, the identity of this fern was always a bit suspect, as noted by the question mark next to the species name on Rod Hill's former web site (Treeferns Down Under). We have re-examined this fern, and based on scale morphology and chloroplast DNA sequence analysis, conclude that it is actually a variant form of Sphaeropteris cooperi (F.V.Muell.) R.M.Tryon, rather than S. tomentosissima.

TABLE 1. Voucher information and GenBank accession numbers for tree ferns examined.

Species	Provenance	ID number	GenBank accession and reference
S. tomentosissima	Papua New Guinea	UC640117	
S. tomentosissima	Papua New Guinea	Conant 4581 (LSC)	Korall et al., 2006 <i>atpA</i> - AM176460 Korall et al., 2007 <i>rbcL</i> – AM177352 <i>trnL</i> intron – AM410304
S. cooperi	Highland Lace "Cultivated" Australia	Yansura 1 (UC)	<i>rbcL</i> – JN106035 <i>atpA</i> - JN106039 <i>trnL</i> intron - JN106036
S. cooperi	"Cultivated" Australia	Yansura 2 (UC)	<i>rbcL</i> – HM347350 <i>atpA</i> – JF690125 <i>trnL</i> intron – JF742607
S. cooperi	Flecker Botanical Gardens, Cairns Australia	Yansura 3 (UC)	<i>rbcL</i> – JN106038 <i>atpA</i> – JN106040 <i>trnL</i> intron – JN1060367

MATERIALS AND METHODS

The type specimen of *Sphaeropteris tomentosissima* (*Cyathea tomentosis-sima* Copel.) was examined and stipe scales were photographed at high resolution at the University and Jepson Herbaria at the University of California Berkeley (Brass 9116; UC 640117).

Leaf material for isolating chloroplast DNA was obtained from three sources: a cultivated plant of Highland Lace and a cultivated Sphaeropteris cooperi, both from the US (Hoshizaki's and Yansura's gardens); and four S. cooperi plants in the Flecker Botanical Gardens in Cairns, Australia. The latter four plants were carefully checked to be sure they had stipe scales consistent with S. cooperi as the garden had one plant labeled Sphaeropteris excelsa (Endlicher) R.M. Tryon, which could be confused with S. cooperi except for the scale differences. DNA was extracted from leaf material using the DNeasy Plant Mini Kit from QIAGEN (Valencia, California, USA), and the purified DNA was then used as a template to amplify three plastid loci (rbcL, atpA, trnL intron) using the polymerase chain reaction (PCR). The reaction was carried out with the appropriate set of primers and Cloned Pfu DNA polymerase from New England Biolabs (Ipswich, MA, USA) according to manufacturer's protocols. The PCR products were purified using the MinElute Reaction Cleanup kit from QIAGEN and then subjected to DNA sequencing on an ABI3730xl DNA Analyzer. All sequences (the four plants from the Flecker Botanical Garden had one common sequence) were deposited in GenBank (Table 1).

The beginning of the *rbcL* gene and the *atpB-rbcL* spacer were amplified with the primers atpBR or atpBR1 and RBCL158R, the middle of the *rbcL* gene with primers brun1 and brun2, and the 3' end as well as the *rbcL-accD* spacer with primers RBCL1187F and ACCD887R. The *atpA* gene was amplified by

TABLE 2.	Primers used	d in amplification and	l sequencing. F	7 = forward;	R = reverse;	S = sequencing.
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Loci primer	Usage	Sequence (5'-3')	Reference This study	
rbcL atpBR	F	TGAGCTTTGGCAATATTATTG		
rbcL atpBR1	F	TAATCTCTTGACCCGCTGGGTTAC	This study	
rbcL RBCL158R	RS	AAGATTCCGCAGCTACTGCAGCTCC	Pryer, 2004	
rbcL brun1	FS	CATTACCTTCACGAGCAAGGTCACGG	This study	
rbcL RBCL1187F	FS	GGAACYTTGGGACATCCTTGG	Korall,2007	
rbcL ACCD887R	R	TTATCACABCGMGCCCATAATCC	Korall,2007	
rbcL rbcf1	S	CCAAAATTGGGGCTTATCTGCT	This study	
rbcL rbcf2	S	CTAGCTTGGCCTTCTATTGCCG	This study	
atpA ESATPF415F	FS	CARGTTCGACAGCAAGTYTCTCG	Schuettpelz,2006	
atpA ESTRNR46F	RS	GTATAGGTTCRARTCCTATTGGACG	Schuettpelz,2006	
atpA atpAf	S	GACAGACTGGTAAAACAGCAGTAG	This study	
atpA atpAr	S	TTGCCGGTCGAATGCCAGCATTAA	This study	
trnL trn1	RS	ATTTGAACTGGTGACACGAGGATT	This study	
trnL trn2	FS	CGAAATCGGTAGACGCTACGGACT	This study	
trnL trn5	S	CTACCCTGTTCTGTTGGGGGAT	This study	
trnL trn6	S	TCGACGGGGGGCTATTCCAACG	This study	
trnL trn9	S	TCGAGTCTCTGTACCTATC	This study	

PCR using the primers ESATPF415F and ESTRNR46F, and the *trnL* intron and flanking sequences were amplified with the primers trn1 and trn2. All primers used for PCR amplification and sequencing are listed in Table 2.

RESULTS

Our first indication that this tree fern might be misidentified was based on the stipe scales, which did not match the published description for *Sphaeropteris tomentosissima* (Holttum, 1963). Comparison with the type specimen reaffirmed that the two plants were very different in scales, leaf, and other details (Fig. 1 and 2A–C). Surprisingly, however, the stipe scales on Highland Lace closely match those of the commonly cultivated Australian tree

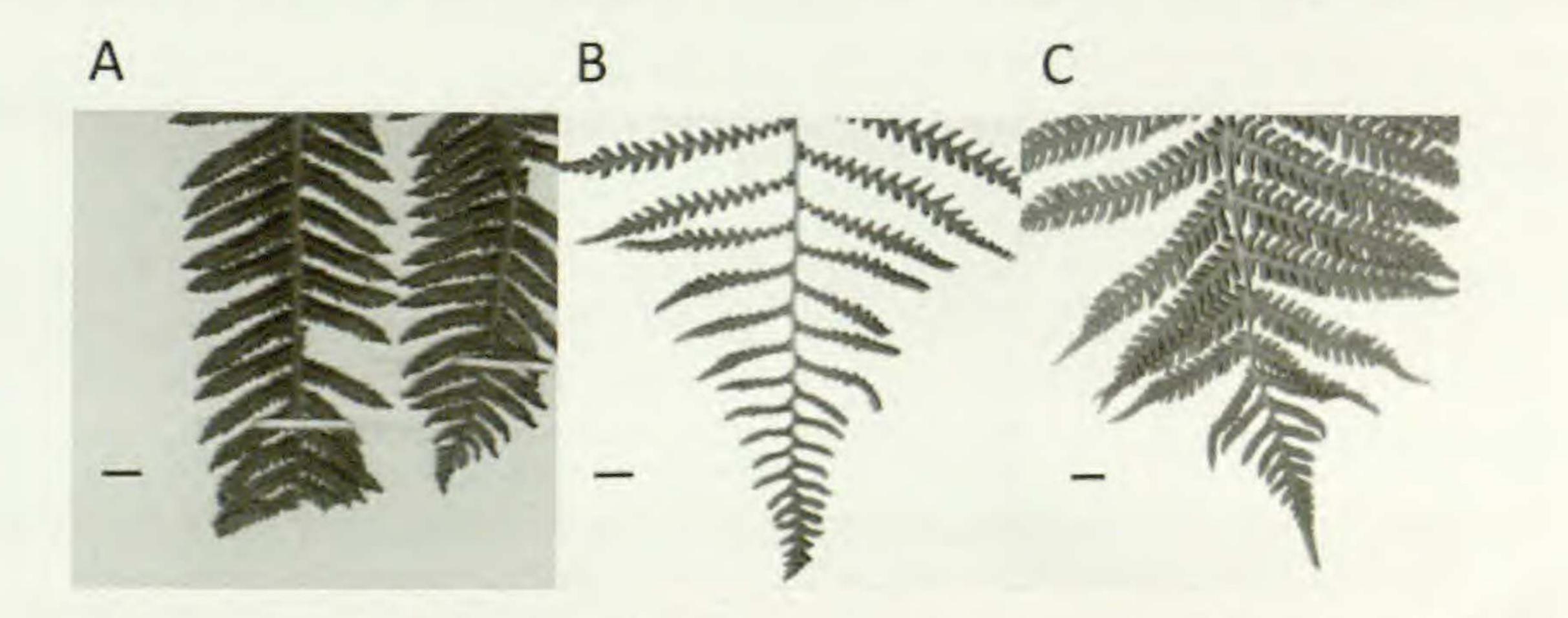


FIG. 1. A comparison of the pinnae. A: Sphaeropteris tomentosissima; B: Highland Lace; C: Sphaeropteris cooperi. The bars represent 1 cm.



A B C

FIG. 2. A comparison of the stipe scales of *Sphaeropteris tomentosissima*, Highland Lace, and *Sphaeropteris cooperi*. A: Scales of *S. tomentosissima* are brown and twisted. B and C: Highland Lace and *S. cooperi* respectively both have large pale scales with dark setae on the edge as well as smaller narrow dark-red scales. The bars represent 1 cm.

fern *Sphaeropteris cooperi* despite their distinct differences in pinnae and leaf shape (Fig. 1).

The scales of Highland Lace were compared to Sphaeropteris cooperi and Sphaeropteris tomentosissima. The S. tomentosissma stipe scales are brown, twisted, and have edges bearing setae of the same color as the scales. In contrast, the broader stipe scales of Highland Lace and S. cooperi are whitish to light tan, with their margins usually bearing a very narrow row of dark reddish brown marginal cells and setae of the same color. Additionally, Highland Lace and S. cooperi have small narrow dark reddish brown scales on the stipe, which are absent on S. tomentosissima (Fig. 2A–C). Also particularly noticeable on S. tomentosissima are the very dense mats of small woolly scales on the abaxial side of all rachises (Fig. 3A–C), which are not present on S. cooperi or Highland Lace. The comparison of scales alone is highly suggestive that Highland Lace is much more closely related to S. cooperi than to S. tomentosissima.

In order to ascertain if Highland Lace is possibly Sphaeropteris cooperi and to further rule out Sphaeropteris tomentosissima, we compared rbcL DNA



FIG. 3. A comparison of the costae of *Sphaeropteris tomentosissima*, Highland Lace and *Sphaeropteris cooperi*. A: *S. tomentosissima* has a dense mat of small wooly scales on the abaxial side. B and C: Highland Lace and *S. cooperi* respectively do not have the dense mat of wooly scales.

Dissimilarity matrix indicating the number of base-pair changes observed for the three TABLE 3. loci, rbcL, atpA and trnL intron. Numbers in parenthesis indicate the total base pairs compared.

		Highland Lace	Cultivated S. cooperi	Flecker S. cooperi	S. tomentosissima
S. tomentosissima	rbcL	4 (1309)	4 (1309)	4 (1309)	-
	atpA	1 (1514)	1 (1514)	1 (1514)	
	trnL	2 (554)	2 (554)	2 (554)	
Flecker S. cooperi	rbcL	0 (1428)	0 (1428)	-	
	atpA	0 (1521)	0 (1521)	-	
	trnL	0 (554)	0 (554)	-	
Cultivated S. cooperi	rbcL	0 (1428)	-		
	atpA	0 (1521)			
	trnL	0 (554)			
Highland Lace	rbcL	_			
	atpA	-			
	trnL				

sequence data for both species (Newmaster et al., 2006; Korall et al., 2007). The Highland Lace sequence differed from the partial gene sequence of S. tomentosissima in GenBank by four changes over the 1309 base pair (bp) length, further evidence that they were different species (Table 3). This sequence was then searched on GenBank and surprisingly the top BLAST match for Highland Lace was a sequence from Sphaeropteris excelsa rather than the expected S. cooperi. The 1309 bp sequence of S. excelsa (AM410213)

was identical to that portion in Highland Lace, while the S. cooperi sequence (SCU05944) differed by four changes over 1320 bp.

This *rbcL* sequence comparison seemed to indicate that Highland Lace was closer to S. excelsa than to either S. cooperi or S. tomentosissima. However, the scales on Highland Lace did not match this conclusion. Highland Lace and S. cooperi have both broad pale scales as well as small narrow dark red scales on its stipe, while S. excelsa has only broad pale scales. Additionally, Highland Lace (and S. cooperi) has small narrow dark red scales on its costa and costule while S. excelsa has a mat of whitish scales and hairs (Hoshizaki and Yansura, 2005). Since the scales on Highland Lace matched those of S. cooperi rather than S. excelsa, we decided to obtain additional S. cooperi rbcL sequence data from a cultivated plant and from four plants from the Flecker Botanical Gardens (Table 1). All five sequences were identical over the complete *rbcL* gene of 1428 bp, and these were exact matches for the Highland Lace gene.

In order to further confirm the identity of Highland Lace, the chloroplast atpA gene sequence (Schuettpetz et al., 2006) was obtained from this plant, from the cultivated S. cooperi plant and from the four tree ferns in the Flecker Botanical Garden. All six sequences matched perfectly over the complete gene sequence of 1521 bp, while the GenBank partial sequence for Sphaeropteris tomentosissima differed by one change over 1514 bp (Table 3), resulting in one

amino acid change (T213N). There were no reference atpA sequences in GenBank for S. cooperi or S. excelsa.

As a final step, we obtained DNA sequences for the trnL (UAA) intron (Taberlet et al., 2007). Highland Lace perfectly matched that of cultivated S. cooperi and the four Flecker Botanical Gardens specimens over the intron's 554 bp (Table 3), and these sequences also matched exactly the partially overlapping 534 bp of S. cooperi in GenBank (EU554328) and S. excelsa over 525 bp (AM410341). The S. tomentosissima sequence in GenBank differed by two bp over the complete 554 bp overlap (Table 3).

Sphaeropteris excelsa and S. cooperi are closely related (Tryon and Tryon, 1959; Tryon, 1970; Jones and Clemesha, 1981) and share at least partial common rbcL and trnL (UAA) intron DNA sequences. These S. excelsa sequences were subsequently reconfirmed using leaf material from a cultivated plant (Hoshizaki and Yansura, 2005). The phylogenetic relationship between these species is unknown, but less conserved non-coding (Shaw et al., 2005; Kress and Erickson, 2007) or nuclear sequences (Sang, 2002) could resolve this question.

DISCUSSION

The identification of tree ferns is especially difficult when the country of origin is not known (Pryer et al., 2010). While Australia has only eleven native species (Jones and Clemesha, 1980), the possibility of non-native spore arriving from nearby New Guinea or from the collections of tree fern enthusiasts within the country is certainly reasonable. The unique appearance of Highland Lace, in particular the reduced pinnules, almost certainly led Rod Hill to identify it as the non-Australian species S. tomentosissima. Upon reexamining Highland Lace, the traditional use of stipe scales for tree fern identification suggested that this identity was incorrect. The more recently developed approach of using chloroplast or nuclear DNA sequences as barcodes for species identification (Kress et al., 2005; Chase et al., 2005; CBOL Plant Working Group, 2009) has been shown to complement traditional analyses based on morphological characters. While DNA sequence analysis is becoming a more widely used tool for this purpose, the public database is still somewhat limited in terms of species coverage. There are only about 150 rbcL sequences from Sphaeropteris, Cyathea and Alsophila in GenBank, while worldwide there are over 600 Cyatheaceae tree fern species (Large and Bragins, 2004). However, an enlarged DNA database will eventually provide a more robust system. The confirmation that Highland Lace is S. cooperi required the use of both morphological characters and DNA sequence analysis. The early study of stipe scales showed that Highland Lace was not Sphaeropteris tomentosissima, but it did not demonstrate that it was S. cooperi. To do so was more tenuous considering that there are approximately 120 Sphaeropteris species worldwide (Large and Braggins, 2004), many with similar scale morphologies.

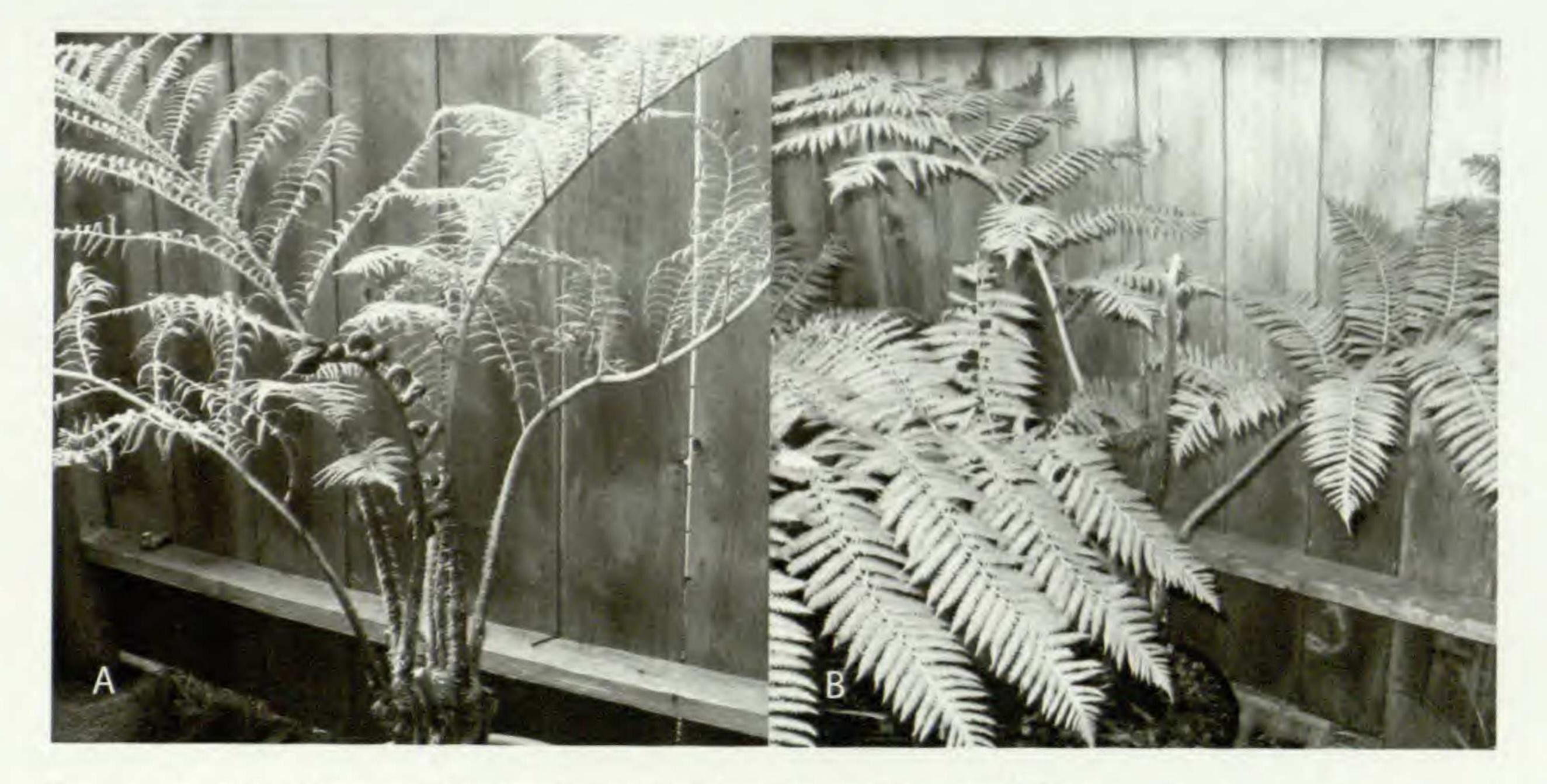


FIG. 4. An overall view of the tree ferns Highland Lace (A) and "Wild-type" Sphaeropteris cooperi (B) showing the significant differences in their general appearance.

Our first DNA sequence analysis based on rbcL confirmed that Highland Lace was not S. tomentosissima, but the effort to determine if it was related to S. cooperi resulted in the discovery of a GenBank voucher that was misidentified (see Results for details). As a result, new reference sequences were made for S. cooperi, which all proved identical to the Highland Lace sequence. Further DNA sequence analysis based on the chloroplast atpA gene and the trnL (UAA) intron also confirmed that Highland Lace is S. cooperi (Table 3). As a practical way to identify a tree fern species, DNA barcoding is an important tool, but with the limited data available, it cannot be used exclusively. The initial Highland Lace rbcL sequence quickly showed that this tree fern was not S. tomentosissima. However, given the sequences that currently exist in GenBank, DNA barcoding could not distinguish whether S. excelsa or S. cooperi was the correct species. Morphologically specific features, particularly the leaf scales in tree ferns, still play an important role in fern identification. The use of morphological characters that initiated this investigation later led to the discovery of the error in the database and its subsequent correction, and scale characteristics ultimately allowed us to choose S. cooperi as the correct species. The interplay of these two methods was important throughout this study.

At first glance, it is difficult to think that Highland Lace and S. cooperi are actually the same species because their general appearances are so strikingly different (Fig. 4). Sphaeropteris cooperi is native to eastern coastal Australia and is known to be variable in form (producing cultivars including Brentwood, Robusta, Allyn Lace, and Allyn Kiest). Most of these variants, however, are quite modest compared to what is observed in Highland Lace with its conspicuously contracted, recurved margins and the reduced size of the

pinnules. Sphaeropteris cooperi shares this ability to produce multiple variants with a limited number of other ferns. Species such as Athyrium filix-femina (L.) Roth and Polystichum setiferum (Forssk.) Moore ex Woynar are also known to produce many variants that have contracted or reduced blade surfaces, recurved margins, and smaller dimensions, plus many more deviations from the typical shape (Rickard, 2000; Hoshizaki and Moran, 2001). A search of the literature suggests that this unusual tree fern may have been reported earlier. A description of Sphaeropteris cooperi (in Flora of Australia, 1998) mentions the existence of an unnamed narrow pinnule variant:

"An occasionally cultivated form of Cyathea (Sphaeropteris) cooperi from central and northern Queensland has narrow recurved abaxially glaucous pinnule lobes, with the majority of rhizome and stipe scales lacking any brown coloration. The sori in this form are commonly restricted to the basal part of each pinnule, at least on younger plants. The Victorian collection may be an isolated accidental occurrence rather than a sample from a naturalized population."

This is possibly the same plant as Highland Lace. However, in the Flora of Australia description, the stem and stipe scales of this fern are said to lack any brown coloration while the Highland Lace specimens in the US have dark redbrown margins, bearing setae. The sori on Highland Lace also may extend well beyond the basal part of each pinnule to near the tip in the US specimens. If Rod Hill's website is correct concerning the origin of this unusual fern in a spore pan (but in New South Wales instead of Queensland), we may consider

this form an accidental occurrence. However it cannot be ruled out that this aberrant plant may also exist in the wild.

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